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LIPID AND SILVER NANOPARTICLES GELS FORMULATION OF TEMPEH EXTRACT

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ABSTRACT

Tempeh extract is used in this study as an active ingredient in lipid nanoparticles and reductant in silver nanoparticles because tempeh is an authentic Indonesian food ingredient and is known to have the main content of isoflavones. Gel preparations were chosen to increase the acceptability and stability of lipid and silver nanoparticles. This research aim is to formulate lipid nanoparticle gel formulations with tempeh extract as active substances and silver nanoparticle gel formulations with tempeh extract as bioreduction. Lipid nanoparticles were made from soy lecithin phospholipids by heating at 60°C and sonication method for 30 minutes then the tempeh extract was added just before sonication. Silver nanoparticles were made by adding tempeh extract to AgNO₃ solution at 90°C for 30 minutes. The average particle size of tempeh extract lipid nanoparticles was 130.03 nm and silver nanoparticle was 94.76 nm. The average viscosity of tempeh extract lipid nanoparticles gel was 4.02 d.Pa.s and silver nanoparticles was 4.22 d.Pa.s. The average spreadability of tempeh extract lipid nanoparticles gel was 4.37 cm and silver nanoparticles is 4.05 cm. The average pH value of tempeh extract lipid nanoparticles was 7.70 and silver nanoparticles was 7.33.

Keywords: gel; lipid nanoparticles; particle size; silver nanoparticles; tempeh extract

INTRODUCTION

Nanoparticles are one of the technologies developed to increase the effectiveness of drug delivery (Latarissa 2017). Nanoparticles have the advantage to penetrate the space between cells and it able to increase the surface area contact. Nano-sized particles have unique physical properties because they can be combined with a variety of technologies. They are expected to produce a more effective drug delivery system (Martien *et al.* 2012). Nanoparticles can be made with specific colloidal formation systems, and one example is liposomes that are made using soy lecithin (Dwiastuti, Noegrohati, and Istyastono 2016). Another method for preparation of nanoparticles is to use metals then reduced with specific materials to form nanoparticles, one example is silver nanoparticles using AgNO₃ solution added with specific reducing agents (Sileikaite *et al.* 2006).

Lipid nanoparticles are made through the formation of soy lecithin phospholipid nanoliposomes by heating and sonication methods (Dwiastuti, Noegrohati, Istyastono, *et al.* 2016). Soy lecithin contains unsaturated fatty acids. It has excellent penetration in the skin and high compatibility in the body (Dwiastuti, Noegrohati, Istyastono, *et al.* 2016). Lipid nanoparticles can combine lipophilic and hydrophilic properties in preparations (Dwinna 2010). Silver nanoparticles are produced through a method of mixing AgNO₃ solution (Tatang Wahyuni, Doni Sugiyama 2011) and specific bioreduction (Muliadi *et al.* 2015). Bioreduction are extracts of natural substances that can act as reductant (Jain D *et al.* 2009). The success of silver nanoparticle formation can be known shortly after manufacture by measuring the maximum wavelength using UV-Vis spectrophotometry (Jain, Arora, *et al.* 2009).

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Tempeh extract on lipid nanoparticles is used as an active substance, while tempeh extract on silver nanoparticles is used as a bioreduction. Tempeh extract is known to have the main content of isoflavones derived from flavonoid compounds that function as wound healers (Park *et al.* 2011). In this research, lipid nanoparticles and silver nanoparticles were formulated to review the physical properties between the two preparations. Lipid nanoparticles were developed as topical preparations because they have good penetration ability (zur Mühlen *et al.* 1998) (Jafar *et al.* 2015). Silver nanoparticles were developed as topical preparations because they have the antibacterial ability (Ariyanta 2014). It can be developed in preparations for wound healing preparations (Ariyanta 2014) and anti-acne (Septiyarin 2017). The development of these two preparations needs to be reviewed for particle size and physical properties as seen from the parameters of viscosity, dispersion, and pH. This preparation is expected to be a choice of drug dosage forms, especially topical preparations for various expected pharmacological effects, for example: wound healing and anti-acne preparations. Therefore, this study aims to formulate lipid nanoparticle gel formulations with tempeh extract as active substances and silver nanoparticle gel formulations with tempeh extract as bioreduction with a review of physical properties and particle size.

METHODS

Materials

The material used in this study were: soybean lecithin (Sigma-Aldrich), distilled water, tempeh with three days fermentation under the brand name "Muchlar", AgNO₃, Carbopol, Propylenglycol, Triethanolamin, and Glycerin are obtained from "Bratachem".

Instrumentation

Instruments used in this study are, particle size analyzer (HORIBA Scientific, Japan), Spectrophotometer UV-VIS (Shimadzu, JAPAN), pH meter, and Viscosimeter Rheosys (Model: Merlin VR).

Preparation of Tempeh Extract

The tempeh extract was prepared by tempeh with three days fermentation under the brand name "Muchlar". Tempeh was cut 5 cm long and 6.5 cm wide and 1 cm thick. Tempeh extract was made with the ratio of tempeh and aquadest which is 1: 2. Three hundred grams of tempeh was added into 600 mL of distilled water, then heated to a temperature of 90°C. Maintained the temperature remained 90°C for 30 minutes then the extract cooled to a temperature 30°C then filter with filter paper.

Preparation of Lipid Nanoparticles of Tempeh Extract as the Active Substances

Lipid nanoparticles were made by weighing soybean lecithin by 12 grams and then minimized by mortar and stamper. The refined soy lecithin was then homogeneously dispersed in 200 mL of aquabidest at 60°C. The soy lecithin dispersion was then blended at high speed for sixty seconds. The soy lecithin suspension was maintained at 60°C and then homogenized with ultraturax for one minute on 4 scale. Furthermore, soy lecithin suspension was put in the bath sonicator together with tempeh extract as much as 80 mL. The sonicator bath is set to a temperature of 60°C for 30 minutes (Dwiastuti, Noegrohati, Istyastono, *et al.* 2016).

Preparation of Silver Nanoparticles Using Tempeh Extract as Bioreduction

Silver nanoparticles were made by weighing 0.034 grams of silver nitrate (AgNO₃) in 200 mL aquabidest (1mM) silver nitrate solution. The silver nitrate solution was heated to a temperature of 90°C. Then it was added with tempeh extract 80 mL and kept at 90°C while stirring 600 rpm for 30 minutes (Ramadon and Mun'im 2016; Ariyanta 2014).

Preparation of Gel and Physical Properties Testing

The preparation of this formula began with the swelling of carbopol. It was prepared in 100 mL lipid nanoparticles or 100 mL silver nanoparticles with 3 grams of carbopol for 24 hours. Then, 3 grams of carbopol 3% w/v as much as 50 grams and added TEA to the

mortar and stirred until homogeneous for about 5 minutes. Next, put the mixture of carbopol and TEA into the blender and add propylene glycol and glycerin and mixing or three minutes at low speed.

Table I. Gel Formula of Lipid and Silver Nanoparticles Gel

Ingredients	Formula
R/ Carbopol 3% b/v (gram)	50
Propyleneglycol (gram)	30
Glycerin (gram)	60
Triethanolamin (TEA) (gram)	2,4

Scattering Test. The scatter power test was carried out 24 hours after manufacture by putting one gram of gel and placed in the middle of a large round glass. On top of the gel was placed another round glass and ballast with a total weight of 125 grams then allowed to stand for one minute and note the spread diameter. **Viscosity Test.** The viscosity test was carried out 24 hours after preparing the gel using the Rheosys cone and plate Merlin VR model. **pH test.** The pH test carried out 24 hours after the gel prepared using a pH-meter. The pH test began with putting one gram of gel and then dissolved it in 10 mL aquadest. Furthermore, the pH meter inserted into the aquadest and then put into a gel then the pH meter will show the pH value.

Wavelength of Silver Nanoparticles.

Measurement of the maximum wavelength is one of the initial steps to determine silver nanoparticles. The indicator of silver nanoparticles is the wavelength with maximum absorbance in the range of 400-450 nm (Ariyanta, 2014; Ayu 2015).

Particles Size of Lipid Nanoparticles and Silver Nanoparticles.

This measurement is done by conducting a DLS particle size analyzer (Horiba SZ 100, Japan).

Data analysis

Particle size data and physical properties test results obtained in this study were then performed statistical tests with the R computational statistical program. The T test

used to find out whether there are significant differences in physical properties results between lipid nanoparticles with silver nanoparticles preparations.

RESULTS AND DISCUSSION

Tempeh extract contains a lot of isoflavones with a function as a wound healing (Danciu *et al.* 2012). Tempeh extract was prepared with water solvent so that the tempeh extract can be used as a bioreduction in the formation of silver nanoparticles. One of the bioreduction requirements in the formation of silver nanoparticles is a water-soluble extract. That is expected to dissolve and react with AgNO₃ solution. While in the addition of lipid nanoparticles, tempeh extract functions as an active substance.

Physical Appearance of Tempeh Extract Lipid Nanoparticles and Tempeh Extract Silver Nanoparticles

The description of lipid nanoparticle was a turbid white color and unique smelled of soy lecithin. The silver nanoparticle preparations had a clear-reddish-brown and unique smelled of tempeh extract. Clear-reddish-brown in the aqueous solution formed from excitation. The reduction of silver ion causes it; there are indicated the formation of silver nanoparticles (Jain, Daima, *et al.* 2009). This physical appearance of difference nanoparticle preparation and gel nanoparticle is presented in Figure 1 and 2.

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The Particle Size of Tempeh Extract Lipid Nanoparticle and Tempeh Extract Silver Nanoparticles

The lipid nanoparticles formation can be known after the Particle Size Analysis (PSA) test have been done. The formation of silver nanoparticles can be recognized immediately by measuring the maximum wavelength using UV Vis spectrophotometer. If the wavelength is between 400 - 450 nm, it means that silver nanoparticles are known (Maharini *et al.* 2017). This is one of the advantages of silver nanoparticles compared to lipid nanoparticles, namely the success of the preparation formulation can be known after manufacture.

In this study, wavelength measurements were made after 24 hours of storage. The average wavelength measurements of silver nanoparticles with three replications after 24 hours of storage at room temperature were obtained 406 nm. The results of these wavelength measurements indicate that silver nanoparticles can be formed the extract of tempeh as bioreduction at a temperature of 90°C and 30 minutes (Sari Purwo Ismaya *et al.* 2017).

PSA test was conducted to determine the size of lipid nanoparticles (Dwiastuti, Noegrohati, Istyastono, *et al.* 2016) and silver nanoparticles. PSA test results are shown in Table II.

The measurement results in Table II showed that the silver nanoparticle formula could produce particle sizes less than 100 nm, while the lipid nanoparticle formulas produce

sizes more than 100 nm. Tempeh Extract in lipid nanoparticle as an active substance make colloidal dispersion could not be form completely so that affect particles size. Tempeh extract in silver nanoparticle will act as bioreductor in silver nitrate and could produce nanoparticle. Analysis with T-test at 95% confidence level obtained p-value of 0.21. Thus the average particle size of tempeh extract lipid nanoparticles was the same as the average particle size of tempeh extract silver nanoparticles and not significantly different. This phenomena could be happen because in silver nanoparticle extract tempeh will initiate reduction reaction of silver nitrate and reduce particle size, but in lipid nanoparticle, particle will be reduce by the reaction of colloidal dispersion from soy lecithin (Dwiastuti, Noegrohati, Istyastono, *et al.* 2016).



Figure 1. Tempeh extract nanoparticle preparation of lipid nanoparticles (a) and silver nanoparticles (b)



Figure 2. Gel nanoparticle of lipid nanoparticle (a) and silver nanoparticle (b)

Table II. Particle Size Analyzer (PSA) result of Lipid and Silver Nanoparticles with Tempeh Extract

Replication	Tempeh Extract Nanoparticle Lipid (nm)	Tempeh Extract Silver Nanoparticles (nm)
Replication 1	129,00	128,10
Replication 2	124,00	87,00
Replication 3	124,20	69,20
Average	130,03 ± 6,41	94,76 ± 30,20

Table III. Viscosity, Spread ability, and pH Value Result of Lipid and Silver Nanoparticles

Parameter	Lipid Nanoparticle	Silver Nanoparticle	<i>p</i> value	Statistical Result
Viscosity (d.Pa.s)	4,02 ± 0,20	4,22 ± 0,33	0,59	Not Significantly Different
Spread ability (cm)	4,37 ± 0,11	4,05 ± 0,02	0,99	Not Significantly Different
pH	7,70 ± 0,10	7,33 ± 0,05	0,98	Not Significantly Different

Physical Properties of Tempeh Extract Lipid Nanoparticles Gel and Tempeh Extract Silver Nanoparticle Gel Preparations

Preparation of tempeh extract lipid nanoparticles and tempeh extract silver nanoparticles were tested for physical properties with parameters including: viscosity, spread ability, and pH value. Physical test results of lipid nanoparticle gel and silver nanoparticle gel preparations showed physical properties test results with pH parameters. The results of the physical properties test were followed by an analysis of the T-test with a 95% confidence level to see differences in physical properties of the two preparations.

The results of the viscosity testing (Table III) after 24 hours of preparation of lipid nanoparticles and tempeh extracts of silver nanoparticles indicated no different results. This result is strengthened by the results of statistical tests using the T-test. The analytical results showed *p*-value is 0,59 so that it can be said that the viscosity of the two preparations that are not significantly different. Viscosity is influenced by the carbopol composition, because carbopol acts as gelling agent that will form gel-forming matrix (Maheswara 2008). The carbopol composition of Lipid Nanoparticle Gel and Silver Nanoparticle Gel have same composition, thus the result of the viscosity testing are not significantly different

The similar analysis results were also found in the spread ability (Table III) and pH response of lipid nanoparticle gel and tempeh extract silver nanoparticles. Statistical tests with the T-test obtained that the *p* value of the spread ability test is 0.99 and the *p*-value of the pH test is 0.98. It can be explained that the spread ability and pH of the preparations resulting from the formulation of lipid nanoparticles and silver nanoparticles of tempeh extract have no significantly different results. This result can be obtained because the amount of gelling agent and humectant used for the preparation of gel nanoparticle lipid and silver nanoparticle gel preparations uses the same amount. The physical properties of gel preparation are influenced by the gelling agent and humectants used in the formulation. Carbopol act as gelling agent and Propylene glycol act as humectant. Gelling agent will form gel-forming matrix. Humectant will maintain the stability of dosage form by absorbing moisture from the environment and reducing the evaporation of water from the preparation. Because of that, spread ability and viscosity will influence dominantly by carbopol and propylene glycol will influence the stability of dosage form (Maheswara 2008).

CONCLUSIONS

Lipid and silver nanoparticles of tempeh extract can be formulated and the average

particle size of lipid nanoparticles was 130.03 nm and silver nanoparticle was 94.76 nm. The average viscosity of lipid nanoparticles gel was 4.02 d.Pa.s and silver nanoparticles was 4.22 d.Pa.s.. The average spreadability of lipid nanoparticles gel was 4.37 cm and silver nanoparticles is 4.05 cm. The average pH value of tempeh extract lipid nanoparticles was 7.70 and silver nanoparticles was 7.33.

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